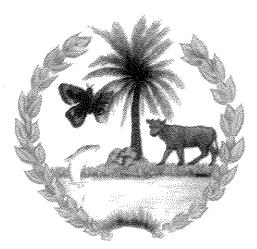
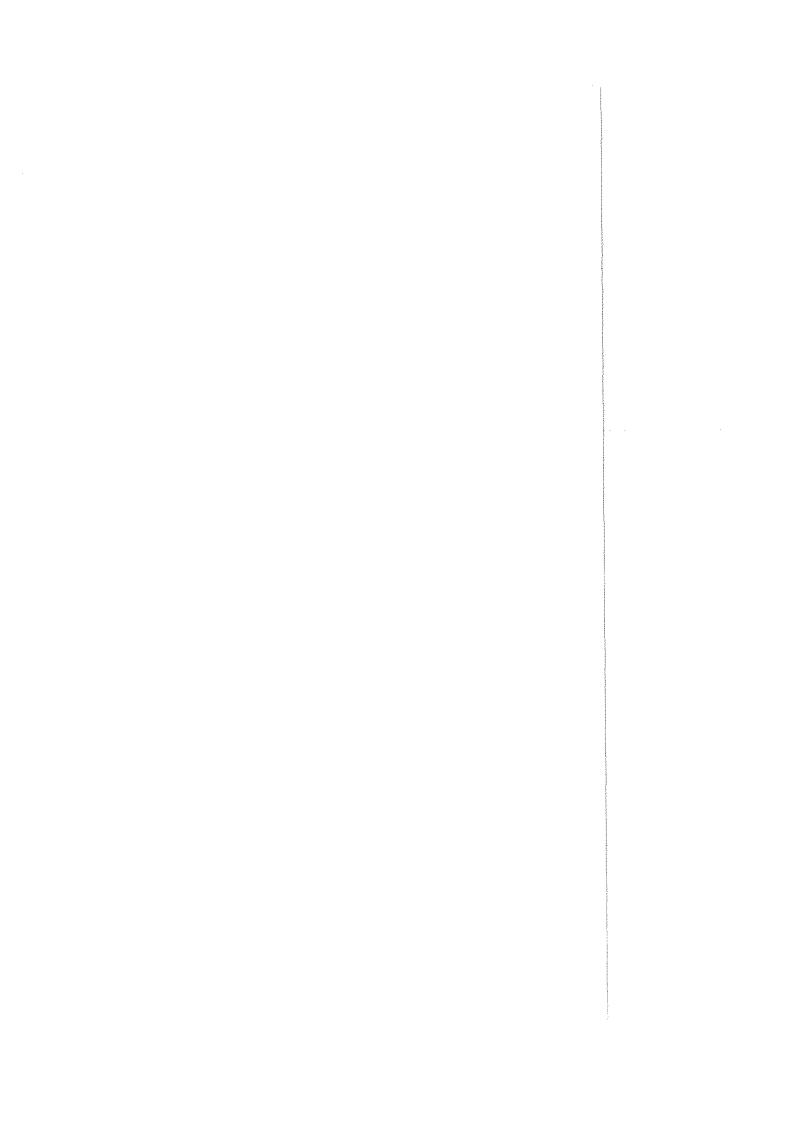
## مجلة إتحاد البيولوجيين العرب القاهرة

# (A) علم الجيوان



http://www.egsz.org Email: info@egsz.org كلية العلوم - جامعة القاهرة



أما مضاد المستقبل -5-HT (Y-25130, 1µM) بقد ثبط قمم الانقباضات في كل من الفنران السليمة والمصابة في جزئي القناة الهضمية، الصائم والقولون، إلا أنه لم يؤثر معنويًا على الفترات الزمنية بين جميع هذه الانقباضات.

نستخلص من هذه النتائج أن (Prostanoid و F-T) يؤثران وظيفيًا على تنظيم الحركة الانقباضية للأمعاء سواءً في الحالات السليمة، أو في حالة الالتهابات الناتجة من الإصابة الطفيلية.

ISSN 1110-5372 Http://www.arabiologists.org Email info@arabiologists.org

مجلة إتحاد البيولوجيين العرب المؤتمر الدولى الرابع عشر ١٥-١٥ ايريل ٢٠٠٧ كلية التربية بالسويس – جامعة قناة السويس – مصر العدد السابع والعشرون (A) علم الحيوان

# تأثير الإصابة بالبلهارسيا على الوظائف الحركية في صائم وقولون الفئران

## فايزه عبده جامعة الملك عبد العزيز - كلية العلوم - قسم علوم الأحياء

تعزى الحركة غير الطبيعية للأمعاء خلال الالتهابات إلى توزيع وإفراز بعض الوسائط الكيميائية مثل البروستانويد Prostanoid وكذلك خامس هيدروكسي التربتامين HT-5، كما تعزى أيضاً إلى انجذاب وتفاعل هذه الوسائط مع المستقبلات المخصصة لها، ولكن ميكانيكية عمل هذه الوسائط والمستقبلات المشاركة أثناء حدوث الالتهاب لا تزال غير معروفة.

أجريت التجارب على مجموعه من الفئران الذكور السويسرية السليمة وكذلك مجموعة من الفئران المصابة بالبلهارسيا (Schistosoma mansoni) لمدة ثمانية أسابيع وبنيت النتائج على أساس مقارنتها بالفئران السليمة. تم إحداث الحركة الانقباضية للقناة الهضمية باستخدام طريقة (Trendelenburg) ، كما حُسبت نتائج متوسط القيم وحللت باستخدام البرامج الإحصائية (paired or un-paired t-tests).

لوحظ زيادة ارتفاعات الانقباضات(Amplitudes) زيادة معنوية في أنسجة الصائم المصابة مقارنة بالسليمة، بينما كان هناك فرقاً معنوياً في الفترات الزمنية (Intervals) بين الانقباضة والأخرى في القولون.

زاد مثبط إنزيمات السيكلواكسيجينيز (نابروكسن) ( naproxen, 10µM من ارتفاع قمم الانقباضات وقلل الفترات الزمنية بينها في كل من صائم الحيوانات السليمة والمصابة، وقد كان هذا التأثير أكثر وضوحًا في الفئران المصابة مقارنة بالسليمة. لكن النابروكسن في القولون قلل من ارتفاع قمم الانقباضات وزاد من الفترات الزمنية بين هذه الانقباضات، وقد كان هذا التأثير أكثر وضوحًا في الفئران المصابة مقارنة بالسليمة.

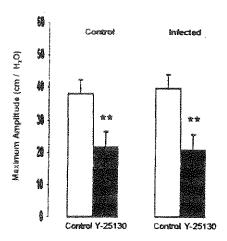


Figure 7A: Effect of 5-HT<sub>3</sub> Receptor Antagonist Y-25130 on MCs in Colon:
Y-25130 significantly inhibited MCs amplitude in both control and infected colon (P<0.01 and P<0.01).

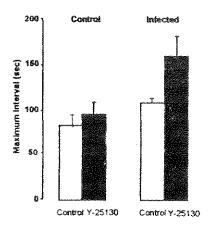


Figure 7B: 5-HT<sub>3</sub> Receptor Antagonist Y-25130 had no effect on MCs intervals.

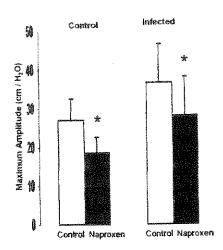


Figure 6A: Effect of COX Inhibitor Naproxen on MCs in the Colon:

In contrast to the findings in the jejunum (Fig. 2A) naproxen significantly attenuated MCs amplitude (P<0.03 and P<0.02).

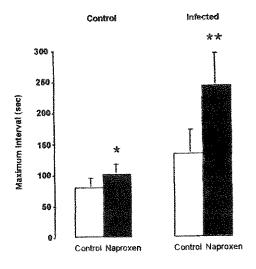


Figure 6B: COX Inhibitor Naproxen increased MCs interval (P<0.02 and P<0.01) in control compared to infected tissue..

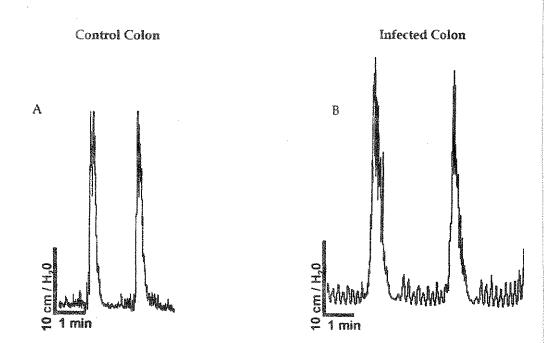


Figure 5: Motor Complex (MCs) in the Colon:

Expanded pressure trace showing the pattern of contractile activity observed in control (A) and infected (B) colon at a distending pressure of  $4-5 \, \text{cm/H}_2\text{O}$ .

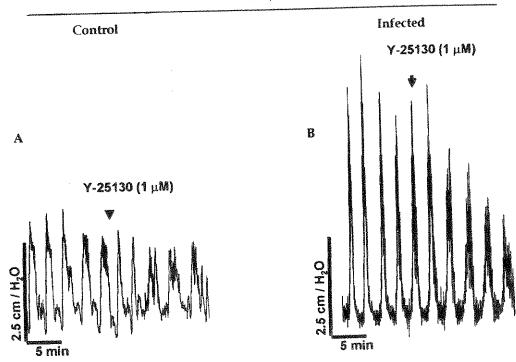
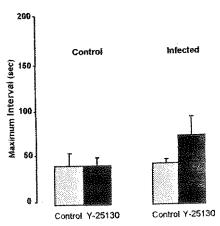


Figure 4 (A+B): Effect of 5-HT<sub>3</sub> Receptor Antagonist Y-25130 on MCs in the jejunum:

(A) Y-25130 significantly inhibited amplitude on MCs in an isolated control jejunum (P<0.05). (B) Y-25130 significantly inhibited MCs amplitude in infected jejunum (P<0.005).



**Figure 4C:** 5-HT<sub>3</sub> Receptor Antagonist Y-25130 had no significant effect on MCs interval

#### Interval

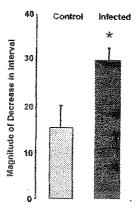


Figure 3: Effect of Naproxen in the Jejunum:

Histogram showing the magnitude of decrease of interval in infected animals (P<0.05).

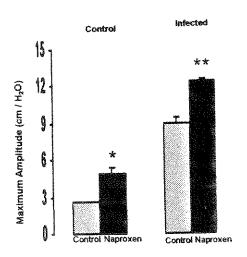


Figure 2A: Effect of COX-2 Inhibitor (Naproxen) on MCs in the jejunum: Histogram depicting the increase of MCs amplitude in control and infected (P<0.05 and P<0.01) jejunum.

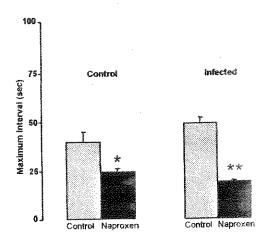


Figure 2B: Effect of COX inhibitor (Naproxen) on MCs in the jejunum
Histogram depicting the decreased of MCs interval evoked by naproxen in control and infected tissues (P< 0.02 and P<0.01, respectively).

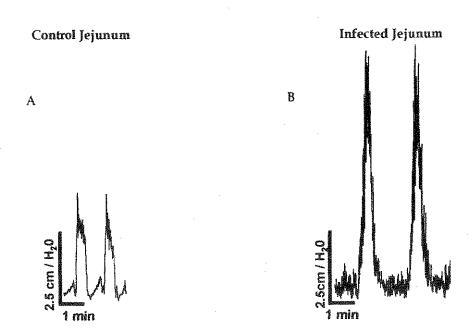


Figure 1: Motor Complexes in the Jejunum:

Expanded pressure trace showing the pattern of contractile activity observed in control and infected jejunum at a distending pressure of 2.5-3 cm/H2O.

- Cyclooxygenase-2 inhibition increases gastric tone and delays gastric emptying in rats. Neurogastroenterol Motil., 19: 225-32.
- SILVA, C.L., MOREL, N., LENZI, H.L. AND NOEL, F. (1998). reactivity to Increased portal hydroxytryptamine of veins from mice infected with Schistosoma mansoni. Comp. Mol. Biochem. Physiol. A Integr. Physiol., 120: 417-23.
- SPILLER, R. (2006). Role of motility in chronic diarrhoea. Neurogastroenterol. Motil., 18: 1045-55.
- SPILLER, R.C. (2002). Role of nerves in enteric infection. Gut., 51: 759-62.
- S., A., HASE, TANAKA, MIYAZAWA, T., OHNO, R. AND TAKEUCHI, K. (2002). Role of cyclooxygenase (COX)-1 COX-2 inhibition and nonsteroidal anti-inflammatory drug-induced intestinal damage relation to various in rats: pathogenic events. J. Pharmacol. Exp. Ther., 303: 1248-54.
- TULADHAR, B.R., KAISAR, M. AND NAYLOR, R.J. (1997).

- Evidence for a 5-HT3 receptor involvement in the facilitation of peristalsis on mucosal application of 5-HT in the guinea pig isolated ileum. Br. J. Pharmacol., 122: 1174-8.
- WADE, P.R., CHEN, J., JAFFE, B., KASSEM, I.S., BLAKELY, R.D. AND GERSHON, M.D. (1996). Localization and function of a 5-HT transporter in crypt epithelia of the gastrointestinal tract. J. Neurosci., 16: 2352-64.
- WARNER, T.D. AND MITCHELL, J.A. (2004). Cyclooxygenases: new forms, new inhibitors, and lessons from the clinic. Faseb. J., 18: 790-804.
- WHELTON, A., WHITE, W.B., BELLO, A.E., PUMA, J.A. AND FORT, J.G. (2002). Effects of celecoxib and rofecoxib on blood pressure and edema in patients > or =65 years of age with systemic hypertension and osteoarthritis. Am. J. Cardiol., 90: 959-63.
- WU, K.K. (1995). Inducible cyclooxygenase and nitric oxide synthase. Adv. Pharmacol., 33: 179-207.

- GRASA, L., ARRUEBO, M.P., PLAZA, M.A. & MURILLO, M.D. (2006). PGE(2) receptors and their intracellular mechanisms in rabbit small intestine. Prostaglandins Other Lipid Mediat., 79: 206-17.
- GRUNDY, D. & SCHEMANN, M. (2004). Serotonin in the gut: pretty when it gets down to the nitty gritty. Neurogastroenterol Motil., 16: 507-9.
- KIYOSUE, M., FUJISAWA, M., KINOSHITA, K., HORI, M. **AND** OZAKI, Η. (2006).Different susceptibilities of spontaneous rhythmicity and myogenic contractility to intestinal muscularis inflammation in the hapteninduced colitis. Neurogastroenterol Motil. 18: 1019-30.
- LORDAL, M. AND HELLSTROM, P.M. (1999). Serotonin stimulates migrating myoelectric complex via 5-HT3-receptors dependent on cholinergic pathways in rat small intestine. Neurogastroenterol Motil., 11: 1-10.
- MANNING. B.P., SHARKEY, K.A. AND MAWE, G.M. (2002). Effects of PGE2 in guinea pig colonic myenteric ganglia. Am. Ĵ. Physiol. Gastrointest. Liver Physiol, 283: G1388-97.
- MIRANDA, A., PELES, S., MCLEAN, P.G. AND SENGUPTA, J.N. (2006). Effects of the 5-HT3 receptor

- antagonist, alosetron, in a rat model of somatic and visceral hyperalgesia. **Pain**, 126: 54-63.
- MOREELS, T.G., DE MAN, J.G., BOGERS, J.J., DE WINTER, B.Y., VROLIX, G., HERMAN, A.G., VAN MARCK, E.A. & PELCKMANS. P.A. (2001).Effect of Schistosoma mansoniinduced granulomatous inflammation on murine gastrointestinal motility. Am. J. Physiol. Gastrointest. Physiol., 280: G1030-42.
- NARUMIYA, S., SUGIMOTO, Y. AND USHIKUBI, F. (1999). Prostanoid receptors: structures, properties, and functions. Physiol. Rev., 79: 1193-226.
- NEVES, S.M., REZENDE, S.A. AND GOES, A.M. (1999). Nitric oxide-mediated immune complex-induced prostaglandin E(2) production by peripheral mononuclear blood cells humans infected with Schistosoma mansoni. Cell Immunol., 195: 37-42.
- NORTHEY, A., DENIS, CIRINO, M., METTERS, K.M. AND NANTEL. F. (2000).Cellular distribution of prostanoid EP receptors mRNA in the rat gastrointestinal tract. Prostaglandins Other Lipid Mediat., 62: 145-56.
- SANTOS, C.L., MEDEIROS, B.A., PALHETA-JUNIOR, R.C., MACEDO, G.M., NOBRE-E-SOUZA, M.A., TRONCON, L.E., SANTOS, A.A. AND SOUZA, M.H. (2007).

Neurogastroenterol Motil, 12: 431-40.

SPENCER, N.J., BUSH, T.G., SANDERS. WATTERS, N., K.M. & SMITH, T.K. (2001). alosetron Effects of migrating motor spontaneous complexes in murine small and large bowel in vitro. Am. J. Liver Gastrointest. Physiol. Physiol., 281: G974-83

DE JONGE, F., VAN NASSAUW, DE MAN, J.G., WINTER, B.Y., VAN MEIR, F., DEPOORTERE, I., PEETERS, T.L., PELCKMANS, P.A., VAN MARCK, E. & TIMMERMANS, (2003).Effects LP. Schistosoma mansoni infection on somatostatin and somatostatin receptor 2A expression in mouse Neurogastroenterol. ileum. Motil., 15: 149-59.

DE MAN, J.G., CHATTERJEE, S., DE WINTER, B.Y., VROLIX, VAN MARCK, E.A., G., & A.G. HERMAN, (2002).PELCKMANS, P.A. somatostatin on Effect of gastrointestinal contractility Schistosoma mansoni infected mice. Int. J. Parasitol., 32: 1309-20

DE MAN, J.G., MOREELS, T.G., DE WINTER, B.Y., BOGERS, MARCK. E.A., VAN  $J_{\cdot \cdot J_{\cdot \cdot \cdot \cdot}}$ AND HERMAN. A.G. (2001).PELCKMANS, P.A. Disturbance of the prejunctional cholinergic of modulation neurotransmission during chronic granulomatous inflammation of the mouse ileum. Br. J. Pharmacol., 133: 695-707.

DEKKERS, J.A., AKKERMANS, L.M. & KROESE, A.B. (1997). Effects of the inflammatory mediator prostaglandin E2 on myenteric neurons in guinea pig ileum. Am. J. Physiol., 272: G1451-6.

DELVAUX, M., LOUVEL, D., MAMET, J.P., CAMPOS-ORIOLA, R. AND FREXINOS, J. (1998). Effect of alosetron on responses to colonic distension in patients with irritable bowel syndrome. Aliment Pharmacol Ther., 12: 849-55.

EL ZAWAWY, L.A., SAID, D.E., GAAFAR, M.R. & ASHRAM, Y.A. (2006). Effect of Schistosoma mansoni infection on physiological gastrointestinal transit and contractility. J. Egypt. Soc. Parasitol., 36: 1057-70.

FORNAI, M., BLANDIZZI, C., ANTONIOLI, L., COLUCCI, R., BERNARDINI, N., SEGNANI, C., DE PONTI, F. AND DEL TACCA, M. (2006). Differential role of cyclooxygenase 1 and 2 isoforms in the modulation of colonic neuromuscular function in experimental inflammation. J. Pharmacol. Exp. Ther, 317: 938-45.

GALLIGAN, J.J. (2004). 5-hydroxytryptamine, ulcerative colitis, and irritable bowel syndrome: molecular connections. Gastroenterology, 126: 1897-9.

terminals of primary afferent nerves intrinsic to the enteric nervous system which then activate downstream interneuron's and motoneurons in enteric neural circuits mediating peristalsis.

The inhibitory effect of Y-25130 on MCs amplitude was also supported by studies of (Bush et al., 2001; Spiller, 2006) who observed that drugs that alter 5-HT signaling have some therapeutic benefit in the treatment of patients with Irritable bowel syndrome (IBS). However, our results which shows that the effect of 5-HT<sub>3</sub> receptor antagonist was less pronounced in the colon when compared to the jejunum are consistent with the recent findings of Berthoud et al., (2004); Grundy and Schemann, (2004) who showed that the mucosal 5-HT synthesis and reuptake were significantly reduced in pathological conditions such as ulcerative colitis and IBS.

In conclusion, the present study suggests that infection with *S. mansoni* leads to disturbance of GI motility. The different effect of infection on amplitude and interval of MCs possibly reflects different actions on the neuromuscular apparatus and the enteric reflex circuits that control the pattern of contractile activity.

#### ACKNOWLEDGMENT

I would like to thank Professor David Grundy and Professor Osama Tayeb for their guidance and consultation. This work is supported by Institute of Research and Consultation, Saudi Arabia.

#### REFERENCES

ABDU, F., G.A., HICKS. HENNIG, G., ALLEN, J.P. & GRUNDY. D. (2002).Somatostatin sst(2) receptors inhibit peristalsis in the rat and mouse jejunum. Am J Physiol Gastrointest Liver Physiol, 282: G624-33.

BERTHOUD, H.R., BLACKSHAW, L.A., BROOKES, S.J. & GRUNDY, D. (2004). Neuroanatomy of extrinsic afferents supplying the gastrointestinal tract. Neurogastroenterol Motil, 16 Suppl 1, 28-33.

BERTRAND, P.P. (2006). Realtime measurement of serotonin release and motility in guinea pig ileum. J. Physiol., 577: 689-704.

BERTRAND, P.P., KUNZE, W.A., FURNESS, J.B. & BORNSTEIN, J.C. (2000). The terminals of myenteric intrinsic primary afferent neurons of the guinea-pig ileum are excited by 5-hydroxytryptamine acting at 5-hydroxytryptamine-3 receptors. Neuroscience., 101: 459-69.

BOGERS, J., MOREELS, T., DE MAN, J., VROLIX, G., JACOBS, W., PELCKMANS, P. & VAN MARCK, E. (2000). Schistosoma mansoni infection causing diffuse enteric inflammation and damage of the enteric nervous system in the mouse small intestine.

the involvement of COX inhibitor displayed different action of this inhibitor on jejunum and colon.

Pattern of contractile activity control and both infected jejunum and colon indicated that intraluminal distension in 8-wk post infection produced an increase of amplitude and colonic jejunual MCs intervals. Thus, obviously receptors-mediated different contractions were involved during inflammation. This was consistent with De Man et al., (2001) results who found that infection with Smansoni disturbed the cholinergic enteric neurotransmission and may contributed to the intestinal motility disturbances. The outcome of the study were also supported by the hypothesis which declares that the clinical symptoms of intestinal Schistosomiasis may be mediated by a dysfunction of GI motor function (De Jonge et al., 2003; El Zawawy et al., 2006).

In the present work, Cyclonaproxen inhibitor oxygenase (10µM) increased the frequency of MCs in the jejunum and inhibited MCs in the colon of control and infected mice that implied to the role of endogenous PG on the activation of different receptor subtypes in various regions of GI tract. This conclusion was similar to Narumiya et al., (1999); Grasa et al., (2006); Santos et al., (2007) that demonstrated whom receptors were found in different regions of GI tract and functionally divided into contractile and relaxant mediate diverse receptors that

PG. Again effect of this supported assumption was Dekkers et al., (1997) who reported that PG evoked contraction in longitudinal muscle and relaxation in circular muscle entail to the of different involvement receptors and may indicate that endogenous PG were an essential component for the maintenance of regular contractile activity inflammation and under normal conditions.

It is important to mention that the effect of naproxen on MCs activity were greater in infected compared to control animals suggesting that the inhibition of cyclo-oxygenase by COX inhibitors may affect the production of PG release during infection. Our result was similar to the findings of De Man et al., (2002), They stated that infection with S.mansoni disturbs the release of neurotransmitters and mediated modulation of cholinergic enteric neurotransmission.

5-HT3 -receptor antagonist Y-25130 used in this investigation inhibited MCs amplitude of mouse jejunum and colon during basal These results conditions. supported by the hypothesis of Galligan, (2004) who suggested that modulation of 5-HT release from EC cells is critical to normal and abnormal GI function because EC cell are sensory transducers that mechanical stimuli to respond applied to the mucosa causing 5-HT release. The release of 5-HT from EC cells can then act at 5-HT3 and 5-HT4 receptors localized on the

P<0.05). At the mean time there was no effect on MCs amplitude  $(34\pm4 \ \nu s \ 39\pm4.5, \ P>0.05, \ Fig. 5 \ A+B)$ .

### Effect of Naproxen on MCs in the Colon

The effect of COX inhibitor naproxen (10µM) on the colon produced opposite effect to the jejunum. In control and infected colon naproxen repressed MCs frequency. MCs amplitude was significantly in both decreased control (27.19±5.5 18.5±4. VS P<0.05, Fig. 6A) and infected  $(36.77\pm10 \text{ vs } 28.32\pm10, \text{ P}<0.05)$ tissue compared to the observation in jejunum in which naproxen had a higher effect on MCs amplitude. Naproxen in the colon had a pronounced effect on MCs intervals. Such an effect was greater in infected tissue compared to (137,29±37s control. 245.27±49s, P<0.01 and 81.09±14s vs  $105.31\pm14$ s, P<0.05 respectively, Fig. 6B).

#### Effect of 5-HT<sub>3</sub> in the Colon

5-HT<sub>3</sub> receptor antagonist (LuM) inhibited the amplitude in  $(39.93 \pm 4.5)$ control both 21.387±2.5, P<0.01) and 8-wk  $(39.72\pm4.5)$ colon infected 20.55±2.5, P < 0.01Fig. However, the magnitude of the inhibition was less marked in the colon (P>0.05) when compared to jejunum  $(P \le 0.05)$ . **MCs** intervals were not affected by Y-25130 in both control (77.71 $\pm$ 13s vs 92.15±17s, P<0.09) and infected (113.35±5s vs 164.93±20s, P<0.06) tissues. Although there was an increase in MCs intervals in infected tissue, this increase did not reach the significant value (Fig. 7B).

#### DISCUSSION

Intestinal inflammation with *S.mansoni* leads to alteration in GI motility. However, the mechanisms underlying these alterations and the receptor involved in the inflammatory response were not fully determined (De Jonge *et al.*, 2003).

In this study, the pattern of contractile activity in response to intraluminal distension in 8-wk infected mice differs from the response in control mice in both jejunum and colon. In the jejunum, Schistosomiasis induced significant increased in MCs amplitude while it triggered increased in MCs. intervals in 8-wk infected colon. These observations were agreement with De Man et al., (2002); El Zawawy et al., (2006), since S. mansonai infection induced smooth muscle hyper contractility of the small intestine.

Although the pathogenesis of *Schistosomiasis* have been extensively studied, little investigations have been focused on the role of inflammatory mediators such as 5-HT and PG on motor patterns of contractile activity in the small and large bowel wall during infection. Moreover, this study on

compared to control. The activity consisted of periodic increases in intraluminal pressure separated by periods of relative quiescent.

The increase in intraluminal pressure coincided with wave of contraction seen as parallel line propagates from oral to aboral. Base line pressure was about the infected in control and same However, control animals. in jejunum, MCs had a maximum pressure of 2.65±0.25  $cmH_2O$ , separated by intervals of 38.8±10 While in (Fig. -1A). seconds infected jejunum MCs had a maximum pressure of  $8.58 \pm 1$ cmH<sub>2</sub>O and intervals of 52.57±6 seconds (Fig. 1B). The amplitude of **MCs** was increased jejunal tissues from significantly in infected jejunum compared control, whereas there was no significant effect of on MCs intervals.

### Effect of COX-1 Inhibitor on MCs of the Jejunum

In control jejunum the COX  $(10\mu M)$ inhibitor naproxen increased the amplitude (2.46±0.01 vs 4.93±0.5 cmH<sub>2</sub>O, P<0.05) and decrease the intervals of MCs (39.58±5s vs 24.1±2s, P<0.05, Fig. 2B). In infected jejunum, naproxen amplified the amplitude (8.96±0.5 vs 12.36±0.1 cmH<sub>2</sub>O, P<0.01, Fig. 2A) and reduced the MCs intervals (49.06±3s vs 19.01±0.6s, P<0.01, Fig. 2B). Such an effect that was greater in infected tissue compared to controls (P<0.05, Fig. 3).

### Effect of 5-HT<sub>3</sub> Receptor Antagonist in the Jejunum

In control jejunum 5-HT<sub>3</sub> receptor antagonist (Y-25130, 1μM) induced significant inhibition of amplitude (2.93±0.3 vs 1.84±0.03 cmH<sub>2</sub>O, P<0.05, Fig. 4A). Such an effect was greater in infected tissue compared to control (7.57±1 vs 4.26±1 cmH<sub>2</sub>O, P<0.01, Fig. 4B).

The effect of 5-HT<sub>3</sub> receptor antagonist on MCs intervals was different. Although the intervals in control tissue remain unchanged (45.67±15s vs 45.19±11s, P>0.05) there was a tendency for the interval to increase in infected tissue (47.60±5s vs 78.74±20s, P<0.06 Fig. 4C). However, this increase was not significant.

#### Motor Complex in Colon

Intraluminal distension mouse colon revealed a similar pattern of contractile activity in jejunum, whereas the mouse parameters of peristaltic reflex between jejunum and colon were different. In control and infected colon, the intraluminal pressure required to initiate the peristaltic reflex was higher when compared with jejunum. In control tissue, base line pressure was 4-5 cmH<sub>2</sub>O, while maximum pressure was 34±4 cmH<sub>2</sub>O and the interval was 85±8s. Although infected jejunum revealed in **MCs** significant increase amplitude compared to control (P<0.05, Fig. 1B), infected colon showed significant increase in intervals (84.4±8.3s vs 128.55±17s,

(composition in mM: NaCl 117, KCI 4.7, NaHCO<sub>3</sub> 25, CaCI<sub>2</sub> 2.5, MgCl<sub>2</sub> 1.2, NaH<sub>2</sub>PO<sub>4</sub>.2H<sub>2</sub>O 1.2 and D-glucose 11). Tissues prepared as described by Abdu et al., (2002). Two jejunal and colon segments approximately 5 cm in length were prepared from each animal and four in total were mounted horizontally in separate 20 ml perfusion chambers. Tissues were maintained at 37°C, perfused with Krebs solution at a rate of 5ml/min, and allowed to equilibrate at least 30 min before experiments started. Motor complex (MCs) of jejunual and colon in infected and uninfected mice were monitored and analyzed by using (Neurolog/NL 900D, Digitimer Ltd. Hertfordshire, England) to record contractile activity as changes in intraluminal pressure under volumetric conditions to compare regional differences and their responsiveness to blockade of inflammatory mediators including prostanoids and 5-HT.

#### Experimental Protocol

Isolated jejunal and colon segments were distended to 2-3.5 cmH<sub>2</sub>O and 4-5 cmH<sub>2</sub>O. respectively, to evoke (MCs). Only preparations in which regular MCs were maintained were used for experiments. subsequent Drugs (naproxen and 5-HT<sub>2</sub> receptor antagonist Y-25130) were added to the chambers 15 minutes after stopping perfusion and recording continued for a further 20 minutes

before washing out the drugs and re-instating perfusion.

#### Drugs

All the peptides were purchased from Sigma Chemical (USA) and were dissolved in distilled water unless otherwise stated. 5-HT<sub>3</sub> receptor antagonist (Y-25130) was dissolved in saline (0.9% Na Cl). All drugs were stored at -20°C. Freshly diluted aliquots were maintained on ice during the course of the experiments and added to the bath in microlitre volumes.

#### Data Analysis

MCs were measured in terms of their peak amplitude above baseline (cmH2O), while duration and interval between them were expressed in seconds (s). Pretreatment values were taken during the 15 minutes before drug application and the response effect was monitored in the 15 minutes following application. Results are expressed as absolute values ± standard error of mean (S.E.M). Paired data were compared using Student's t-test. Probability of P<0.05 was considered significant.

#### RESULTS

#### MCs in Jejunum

Luminal distension of isolated segments of mouse jejunum evoked a regular pattern of motor activity. There was no difference between the patterns of activity in infected jejunum

The COX-I enzyme is the major isoform expressed in GI tract (Tanaka et al., 2002; Warner and Mitchell, 2004). It is believed that the adverse effect of nonsteroidal anti-inflammatory drugs (NSAIDs) result from the inhibition of this isoform (Northey et al., 2000).

Naproxen is antagonist with some selectivity at COX-1 isoform. It has been shown to control the formation of PG and reduce the inflammation by inhibiting COX-1 isoform (Warner and Mitchell, 2004).

5-HT is a critical mediator of the peristaltic reflex released from EC cells that respond to mechanical stimuli applied to the mucosa. 5-HT the implicated also pathogenesis of intestinal peristalsis released 5-HT locally since contributes to the initiation of motor reflexes and the transduction of nociceptive stimuli (Delvaux et al., 1998). 5-HT<sub>3</sub> and 5-HT<sub>4</sub> receptor ligands have proved to be effective in the treatment of visceral motility and hypersensitivity disorders (Grundy and Schemann, 2004). Pharmacological blockade of dimiraishes 5-HT<sub>3</sub> receptor propulsive motor activity, reduces peristaltic reflex activity (Wade et al., 1996; Tuladhar et al., 1997).

Therefore, the rational of the present study was to compare the patterns of contractile activity in control animals with other animals that have been subjected to an inflammatory insult and to examine the role of 5-HT and prostanoids by

using 5-HT<sub>3</sub> selective receptor antagonists (Y-25130) and COX inhibitor naproxen on the generation of abnormal motor patterns.

### MATERIAL AND METHODS Schistosoma mansoni Infection

The maintenance of the Smansoni life cycle and transcutaneous infection of mice with S. mansoni were carried out according to the methods of Bogers et al., (2000); Moreels et al., (2001). Swiss male mice (age 7-wk) were transcutaneously infected with about 100 S. mansoni cercariae, then loaded with treated water containing 100 infectious cercariae of a Biomphalaria Alexandriana strain of S. mansoni. The cercariae were allowed to penetrate during 30 min after which the water was removed and checked for remaining cercariae. Control and Infected mice were sacrificed by cervical dislocation after 8-wk of infection and the contractile activity of isolated segments from jejunum and investigated. were colon experiments were approved by the Ethic Committee of King Fahad medical research centre (KFMRC).

#### Tissue Preparation

Control and infected animals were stunned by a blow on the head. A mid-line laparotomy was performed and a segment of proximal jejunum and colon was rapidly excised and placed in gassed (95% O<sub>2</sub> and 5%CO<sub>2</sub>) Krebs bicarbonate buffer solution

considerable attention on studying the mechanisms which generate these symptoms particularly the role of enteric neural circuits that contribute to the intestinal inflammation (Kiyosue et al.. 2006). Schistosomiasis are useful in studying inflammation-induced changes in intestinal sensory motor function. It is characterized by gastrointestinal (GI) motilityrelated disorders (De Jonge et al., 2003) due to the distribution or release of chemical mediators such as prostanoids and 5-HT and to the affinity of their specific receptors.

5-hydroxytryptamine (5-HT. cyclo-oxygenase serotonin) and (COX) are known to act as neurogenic mediators within the enteric neural circutes during inflammation (Silva et al., 1998; Neves et al., 1999; Moreels et al., 2001). They exert their effects through interactions with different receptor subtypes on nerves and muscle cells in the small intestine (Lordal and Hellstrom. Miranda et al., 2006). 5-HT has been found in the enterochromaffin cells (EC) of the mucosa of GI tract (Berthoud et al., 2004; Bertrand. 2006). It is also localized in the cell bodies of neurones in the myenteric plexus (Bertrand et al., 2000), nerve fibres distributed in both mventeric and submucous plexuses (Spiller, 2002) as well as in the lamina propria (Bertrand et al., 2000).

COX is the most common therapeutic drug target in GI tract (Warner and Mitchell, 2004; Fornai

et al., 2006). The two COX enzymes, COX-1 and COX-2, were responsible for the production of prostanoids such as PGE2, PGF2 alfa, Prostacycyclin and thrompoxane (Fornai et al., 2006).

Endogenous prostaglandin (PG) have been shown to be important mediators of GI motility in control conditions (Tanaka et al., 2002). As a result, the COX inhibitor naproxen was used to study the role of PG in the S. mansoni infection-induced alterations of contractile activity.

PG are also known to modify motility during inflammation GI(Manning et al., 2002), but the cell surface receptors mediating these prostanoid actions have not been fully characterized. PGE receptors of the EP1, EP2, EP3 and EP4 type, were expressed in the gut of several species including man (Wu, 1995). Northey et al., 2000). Cellular localization studies indicate that the external muscle layers display EP1 EP2 and EP3 receptors and that some EP3 receptors are associated with enteric neurons of the rat and mouse intestine (Whelton et al., 2002).

Pharmacological investigations have shown that circular muscle cells of the guineapig's ileum exhibit EP1 and EP3 receptors, which mediate contraction on activation by agonists, whereas EP2 receptors bring about relaxation (Northey et al., 2000).

I Union Arab Biol Cairo
Http://www.arabiologists.org
Email\_info@arabiologists.org
Vol (27A) Zoology 245-264,April,2007

ISSN 111-5372 14<sup>th</sup> International Conference 15-19 April, 2007 Faculty of Education of Suez,

Rec 25/4/2006

Suez Canal University, Egypt

### EFFECT OF INTESTINAL SCHISTOSOMIASIS ON MOTOR FUNCTION IN THE MOUSE JEJUNUM AND COLON

#### Faiza Abdu

Department of Bioscience, Faculty of Science, King Abdulaziz University, Jeddah, Saudi Arabia

**Key words:** *Schistosoma mansoni*, Prostanoids, 5 Hydroxy Tryptamine, Motility, Jejunum and Colon.

#### **ABSTRACT**

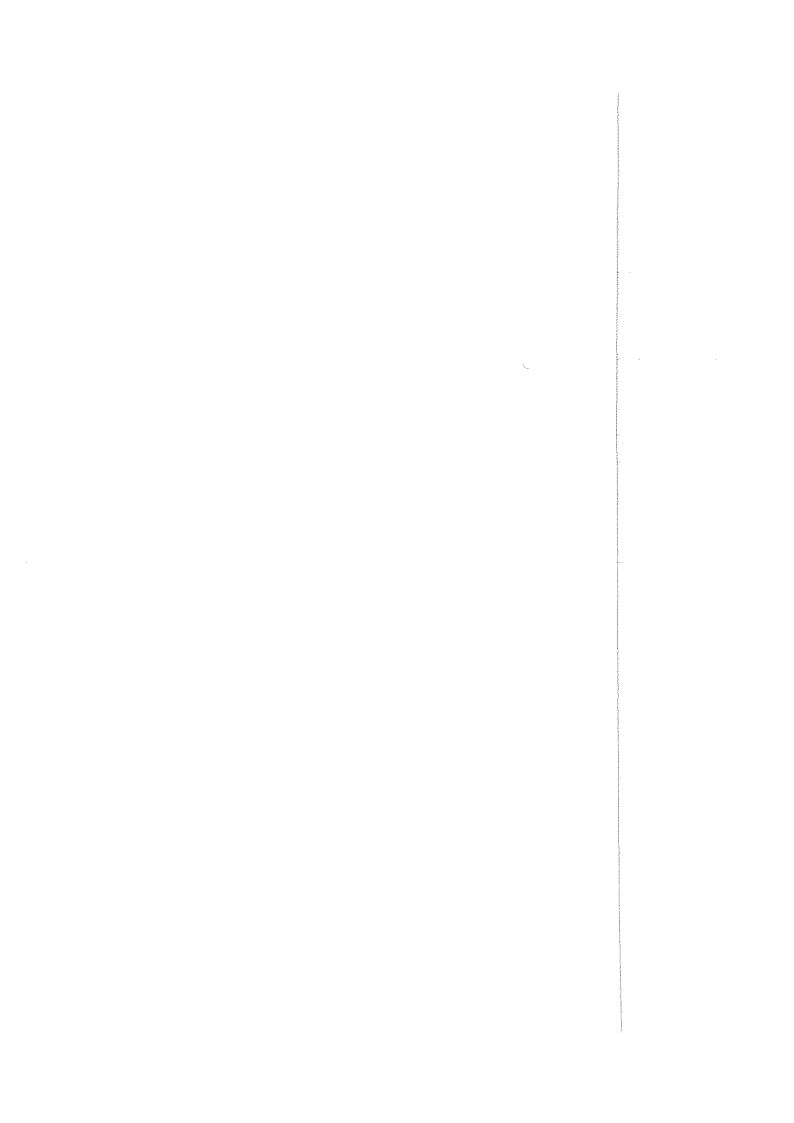
Motor abnormalities during inflammation are related to distribution and/or release of chemical mediators such as prostanoids and 5-Hydroxy Tryptamine (5-HT) in addition to the affinity of their specific receptors. However, the mechanisms of these mediators and the receptors involved are unknown. Experiments were performed on Swiss male mice 8-wk following infection with Schistosoma mansoni compared to uninfected controls. Jejunal and colonic motility was assessed using a Trendelenburg type preparation to study aborally directed motor complexes (MCs). Results showed that the amplitude of jejunal MCs was increased in tissues of infected jejunum compared to control (P<0.05), while in the colon, there was a significant increase in MCs intervals in infected tissue compared to uninfected animals. The cyclo-oxygenase (COX) inhibitor naproxen (10µM) increased the amplitude and shortened the intervals of MCs in both control and infected jejunum; an effect that was greater in infected tissue compared to controls (P<0.05). In contrast, in the colon, naproxen lowered the amplitude and augmented the intervals of MCs. This effect was also higher in tissue of infected animals compared to uninfected control (P<0.01). Although the 5-HTs receptor antagonist, Y-25130 (1µM). inhibited MCs amplitude in both control and infected colon and jejunum, Y-25130 had no effect on MCs intervals.

In conclusion, the present results suggested that infection with *Schistosoma mansoni* lead to alterations in both jejunal and colonic motility. Prostanoids and 5-HT are implicated in both normal motility and in altered function triggered by infection.

#### INTRODUCTION

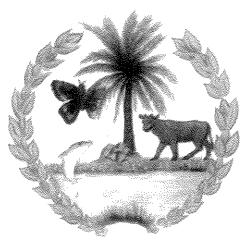
Post-inflammatory changes in intestinal sensory-motor function

are thought to play a major role in the aetiology of irritable bowl symptoms. Currently, there is



### Journal of Union of Arab Biologists Cairo

# (A) Zoology



http://www.egsz.org Email: info@egsz.org Faculty of Science - Cairo University